ISOLATION OF AN ISOMER OF FK-506 FROM FERMENTATION OF <u>Streptomyces</u> tsukubaensis AND ITS CHEMICAL SYNTHESIS FROM FK-506

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Summary: From the fermentation broth of <u>Streptomyces tsukubaensis</u> 9993 an isomer 2a of the immunosuppressant FK-506 la was isolated as a minor metabolite. From NMR- and FAB-MS data follows that in 2a the size of the macrocycle is reduced by two carbon atoms due to a shift of the lactone-acyl from 0-26 to 0-24. The compound was synthesized from FK-506 and proved to be identical with the natural product 2a.

The macrolide FK-506 (1a) is a new powerful immunosuppressant, 1 which has been isolated from the fermentation broth of the soil actinomycete





la (FK-506)

2a (Iso-FK-506)

<u>Streptomyces</u> <u>tsukubaensis</u> 9993 together with several closely-related minor metabolites.² It has a similar mode of action as cyclosporin A, although totally distinct in structure and binding to a different cytosolic receptor.³ Currently FK-506 is in broad clinical evaluation for the prevention and treatment of organ transplant rejection.⁴ We now report the isolation of a new metabolite 2a from <u>S</u>. <u>tsukubaensis</u>, an isomer of FK-506 wherein the lactone-acyl is connected to the oxyfunction in position 24 instead of 26, and its chemical synthesis, starting from FK-506.⁵

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(27.000 1) Fermentation broth
Ethyl acetate extract
         Distribution n-hexane/methanol-water 9:1
                                      hexane layer
Methanol-water layer
Delipidized extract (5.6 kg)
         Sephadex LH 20 chromatography, elution with methanol
Crude FK-506 fraction (980 g)
         Silica gel 60 chromatography (methyl-t.butylether)
Fraction (19.8 g)
                                      FK-506 (420 g)
         Silica gel H (Merck) (methyl-t.butylether)
Fraction (2.6 g)
         Sephadex LH 20 (methanol)
Fraction (1.64 g)
        LiChoprep RP 18 (0.04-0.06 mesh) (water/methanol 20/80)
2a (771 mg)
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Scheme 1: Isolation of the FK 506 isomer 2a

During the isolation of FK-506 from the ethyl acetate extract of the fermentation broth of <u>Streptomyces</u> <u>tsukubaensis</u> according to Scheme 1, we

obtained side fractions which by HPLC analysis on LiChrosorb RP18 showed the presence of a new unidentified component, having a longer retention time than FK-506 (11.70 min vs 10.66 min for FK-506). The compound was isolated and purified by sequential chromatography on silica gel H, Sephadex LH20 and LiChroprep RP18 (Scheme 1) to give an amorphous colourless material with a melting point of $103-105^{\circ}C$.

As with FK-506 the FAB-MS spectrum of the new metabolite is characterized by peaks at 786 (FK-506: $\text{MH}^+\text{-H}_2\text{O}$) and 768 (FK-506: $\text{MH}^+\text{-2H}_2\text{O}$) $^{\text{m}}/\text{z}$ indicating that the compound is an isomer of FK-506. The $^{13}\text{C-NMR}$ spectrum closely resembles the spectrum of FK-506 (comp.table 1) with both, the same total number of C-atom signals and the same characteristic pattern of C(=O)-, C(-O) and C(=C)-signals, being observed. The $^{1}\text{H-NMR}$ spectrum (CDCl₃/CD₃OD 5:1⁶) features a doublet at δ = 3.91 (J = 1.5 Hz). By decoupling experiments it has been assigned to H-26 (FK-506 in CDCl₃: H-26 at δ = 5.33). The H-24 signal is found at δ = 5.29 (FK-506: H-24 at δ = 3.94). Thus the H-24 and H-26 signals exchanged positions indicating an acyl-shift from O-26 to O-24.

These data suggest that the new metabolite has the structure 2a, being an isomer of FK-506 where the size of the macrocycle is reduced by two carbon atoms.



Scheme 2.

To confirm this structure we embarked on the synthesis of 2a starting

Table 1: ¹³C- and ¹H-NMR data of FK-506 (1a), iso-FK-506 (2a), 33-t.-butyldimethylsilyl-22(S)-dihydro-FK-506 (3a) and 33-t.-butyldimethylsilyl-22(S)-dihydro-iso-FK-506 (5a)

	1a ^a		2 a ^b			3a ^a		
Position	¹³ c	1 _H	13 _C	1 _H	13 _C	1 _H	¹³ c	1 _H
1	168.9		168.9		169.1		170.8	
2	56.5		56.4		56.9	4.67	56.3	5.22
3	27.5		26.3		27.3		28.0	1.94
4	21.0		20.7				21.8	2.20
5	24.4		24.9		24.6		24.5	1.80
6	39.1		38.2			4 43	39.4	2.90
8	164 6		162 3		165 0	4.43	166 3	4.48
ğ	196.1		196 5		196 1		105.5	
10	97.0		99.8		96.9		96.2	
11	34.9		33.9				34.6	
11Me	16.1		16.5		16.2		16.3	
12	32.5		33.5		32.4		32.7	1 46
13	73.5		73.5		73.8		73.5	3.35
130Me	56.2							
14	72.7		71.3		72.6	3.63	72.3	3.67
15	75.1		74.0		75.3	3.58	74.9	3.57
150Me	56.7							
16	32,9		31.6		32.8		31.5	0.95 1.40
17	26.1		26.6		26.5		26.1	1.60
17Me	20.3		20.3		20.6		20.7	0.90
18	48.5		50.5		49.3		49.6	1.70 2.16
19	138.8		139.7		136.1		135.9	
19Me	15.8		16.5		15.3		15.3	1.60
20	122.4		123.7		126.3		127.5	4.80
21	52.7		53.3		43.7		43.8	2.71
22	212.4		211.6		71.8		69.7	3.06
23	43.4		41.2	2.65 3.17			34.7	
24	69.8	3.94	71.5	5.29	70.4	3.96	73.2	5.22
25	39.7		37.2	2.02	40.1		40.2	1.80
25Me	9.4		8.2		8.9		8.9	0.96
26	77.3	5.33	77.3	3.91 D 1.5 Hz	77.0	5.31	78.3	3.86
27	13.9		14.1		14.5		12.3	1.60
28	132.2		136.3		132.2		134.5	
29	129.8		127.8		128.6		132.3	5.15
30	34.7		35.0				35.1	2.26
31	34.8		35.1				36.5	0.95
								1.95
32	84.1		84.4		84.1		84.3	2,95
320Me	56.5							
33	73.4		73.4		75.2		75.2	3.40
34	31.2		31.4		33.7		33.9	1.36
								1.88
35	30.5		31.0		30.9		30.8	1.06
							 -	1.60
36	35.1		36.6				32.3	1.84
	105 5				100 -			2.46
3/	133.5		135.1		115 -		137.8	5.72
20	110.3		TT0'9		113.0		112.0	4.92
								4.99

a) In CDCl₃, major conformer; b) In CDCl₃/CDOD₃ 5:1; c) In CDCl₃.



Scheme 3. Reaction conditions: a Im (or DMAP, Py), DMF; b 2%HF, CH₃CN; c pTsOH, toluene.



11a (47%, from 5a)



Scheme 4. Reaction conditions: a NMO, TPAP, 4Å mol sieve, CH₂Cl₂; b 'BuMe₂SiOTf, lutidine; c Me₃SiOTf, lutidine; d IN HCl, CH₃CN.

from FK-506 (1a). We had already earlier detected independently, that a compound of the same structural type as 2a is formed in a reaction of a 22-dihydro-FK-506 derivative: When 33-O-silvlated 22-dihydro-FK-506 (3a) was treated with carbonyldiimidazole to give the cyclic carbonate 4a a byproduct was obtained which was identified as the 22-dihydro-iso-FK-506 derivative 5a (Scheme 2). As in the case of FK-506 and 2a the ¹³C-NMR spectra of 3a and 5a strongly resemble each other (Table 1). In CDCl, only signals of one single conformation (cis-amide: & C-2 56.3 ppm, C-6 39.4 ppm) are seen. In the ¹H-NMR spectrum of 5a as for 2a a doublet appears at $\delta = 3.86$ (J = 6Hz), which is identified as the H-26 signal by decoupling experiments (coupling with H-25, $\delta = 1.80$). The signal of H-24 is located at δ = 5.22 and superimposed on the H-29 signal. A 2D-CH-correlation study confirmed these assignments (data not shown). Since 3a is the major isomer obtained from 33-0-silylated FK-506 (14a) by reduction with triacetoxyborohydride (Scheme 6), the configuration at C-22 in 3a and 5a was assumed to be S. 5,7

Anticipating that the 1,3-acyl shift had been induced by imidazole generated during the carbonylation reaction, we treated 33-O-silvlated 22-dihydro-FK-506 **3a** with imidazole in DMF at 50°C. **5a was formed** in a slow reaction but without major byproducts. Work up after seven days gave a 62% yield of 5a together with unchanged 3a (Scheme 3). The reaction could in principle also be carried out with unprotected dihydro-FK 506 (6a), but the chromatographic isolation of the product proved to be more difficult due to smaller differences in the r_p-values. 4-Dimethylpyridine (DMAP) could be used instead of imidazole. With stronger nonnucleophilic bases like diazabicycloundecane (DBU) additional reactions occurred, on we will report separately.⁸ Interestingly with the dihydro which derivative 3b, the 21-ethyl analogue of 3a, the base catalyzed acyl shift is proceeding even slower: only 47% yield of 5b and 47% recovered starting material were isolated after seven days with DMAP in DMF at 50°C as compared to 70% yield of 5a from 3a under identical conditions.

Selective oxidation of the 22-oxyfunction in 5a to carbonyl failed: Treatment of 5a with N-methyl-morpholine-N-oxide (NMO) and tetrapropylammonium-perruthenate (TPAP) or with periodinane⁹ (Dess-Martin reagent) resulted in preferential oxidation of the 26-oxyfunction. A mixture of the 26-carbonyl- 9a and the 22,26-dicarbonyl-derivative 10a was obtained which could be separated only after silylation of the remaining oxyfunction in 9a to give 11a (Scheme 4). The problem was solved by selective protection of the 26-OH group of 5a, which could be achieved with trimethylsilyltriflate/lutidine in dichloromethane at -70°C, whereas with bis-trimethylsilylacetamide in toluene the 22- and 26-0-silylated products were obtained in about equal amounts. Oxidation of the protected intermediate 12a with NMO/TPAP and subsequent deprotection yielded 2a which proved to be identical with the fermentation product upon TLC and ¹H-NMR comparison.¹⁰

All attempts to isomerize FK-506 (1a) or its 21-ethyl analogue (1b) directly to 2a,b were unsuccessful. FK-506 or its 33-0-tert.butyldimethylsilyl derivative 14a did not rearrange when heated with DMAP in DMF.¹¹ No reaction was observed either with 1b in the presence of tetraisopropoxy-titanium in THF or dichloromethane. When intermolecular methylester transesterification of 14b to the was attempted (tetraisopropoxytitanium/methanol or -/methyl acetate) benzilic acid rearrangement to 15b took place instead, ¹² presumably as result of an attack of methyltitanate at the 9-carbonyl of 1b (Scheme 5).



Scheme 5.

With the 22-dihydro-iso-FK-506 derivatives of **5a**, **b** no further acyl shift to 0-22 or backshift to 0-26 was observed when they were heated with imidazole or DMAP in DMF at 50° C. Under acidic conditions, i.e. p-toluenesulfonic acid in toluene, cyclisation of **5a** or **7a** to the 22,26-epoxide **8a** occurred (Scheme 3). From an ¹H-NMR analysis it follows, that all substituents on the newly formed tetrahydropyran ring of **8a** are in equatorial position. It is assumed that the configuration at C-25 remaines unchanged throughout the series of reactions starting from FK-506. Therefore the configurations at the other positions on the

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Scheme 6. Reaction Conditions: a 'BuMe₂SiCl,Im,DMF; b Me₄NBH(OAc)₃ HOAc/CH₃CN; c LiBH(secBu)₃, THF; d NaBO₃, THF; e 2%HF, CH₃CN. tetrahydropyran moiety follow to be as shown in Scheme 3. This confirms, that the major product from the reduction of FK-506 with the Evans reagent (Scheme 6) is the anti-(S)-diastereomer 3a.^{5,7b}

Furthermore 3a was compared with the product obtained by reduction of FK-506 with Lithiumselectride, for which the 22(S) configuration has been assigned.⁷ In our hands the reaction of FK-506 with Lithiumselectride in THF at -70° C gave mainly a boronic acidester 16a¹³ together with small amounts of 3a. The unsubstituted dihydro derivative was obtained from 16a by treatment with sodiumperborate and was shown to be 3a by TLC- and ¹H-NMR comparison.

EXPERIMENTAL

¹H- and ¹³C-NMR-spectra were recorded at 250 MHz (Bruker WM 250). The concentration was about 20mg in 0.5ml CDCl_3 . The solvent was used as internal standard (δ $\text{CDCl}_3 = 77.0$). SF = 62.9 MHz, SW = 15000 Hz, SI = 32k, AQ = 1.081 sec, PW = 6 sec (corresponding to a 60° pulse), NS = 40000, EM with LB = 1. Proton BB decoupled with DP = 10H. C-H correlation spectra were recorded with the Bruker standard program

XHCORRDC.AU. SW2 = 6000 Hz, SI2 = 4K, SW1 = 700 Hz, NE = 64 a 512 scans, before Fourrier transformation filled with zero's to SI1 = 256 and in both dimensions multiplied with a pi/4 shifted line bell window function.

33-0-tert.Butyldimethylsilyl-22(S)-dihydro-FK-506 (3a)

a) By reduction with tetramethylammonium-triacetoxyborohydride. To 33-O-tert.butyldimethylsilyl-FK-506 (14a)^{1a,14} (23.4g, 25.5 mmol)in 220ml acetonitrile and 80ml acetic acid a solution of tetramethylammonium-triacetoxyborohydride (7.37g, 28 mmol) in 25ml acetonitrile was added dropwise with stirring at -5° C. After 5 h stirring at -5 to 0° C the reaction was quenched by addition of water. The mixture was concentrated in vacuum and the residue partitioned between dichloromethane and water. The organic layer was dried (MgSO₄) and the solvents evaporated. Chromatography of the residue (toluene/ethyl acetate 1:1) gave 15.6g (66%) **3a** along with 2.1g starting material and 1g of a fraction containing starting material and the 22(R)-isomer 17a.

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b) By reduction with Lithiumselectride. A solution of Lithiumselectride (0.57g , 3 mmol) in 3ml THF was added dropwise to 33-0-tert.butyldimethylsilyl-FK-506 (14a) (1.83g ,2 mmol) in 3ml THF stirred at -70°C. After 2 h at -70°C the reaction was quenched by addition of saturated aqueous NH4Cl and the mixture extracted with dichloromethane. The extract yielded after drying (MgSO₄) and evaporation of the solvent upon chromatography of the residue (toluene/ethyl acetate 4:1) 1.67g (48%) 16a and 0.08g 3a. 16a ¹H-NMR (CDCl₂): δ 5.07 (s,br H-26) 4.70 (broad d, J=4Hz, H-2e) 4.45 (d, br J=13Hz, 6e) 4.06 (m, H-22) 3.85 (m, H-24) 3.67 (d, J=8Hz, H-14)3.61 (dd, J=2.5Hz, J=11Hz, H-15) 2.96 (m, H-32). ¹³C-NMR (CDCl₂): δ 196.04 (C-9), 169.16 (C-1), 165.24 (C-8), 137.23 (C-37), 136.15 (C-19), 131.07 (C-28), 129.29 (C-29), 125.68 (C-20), 115.74 (C-38), 96.43 (C-10), 84.13 (C-32), 77.13 (C-26), 75.20 (C-33), 74.86 (C-15), 73.65 (C-13), 72.64 (C-14), 70.37 (C-24), 69.50 + 69.48 (C-22), 58.12 (OMe), 56.98 (OMe), 56.70 (C-2), 56.36 (OMe), 49.62 (C-18), 43.07 (C-21), 40.67 (C-25), 39.41 (C-6), 36.69 (C-23), 36.23 (C-31), 34.90 (C16,C-30), 34.69 (C-11), 33.77 (C-36), 33.31 (C-12,C-34), 30.94 (C-35), 27.69 (C-3), 26.36 (C-17), 26.23 + 26.16 (B-C-CH₂?), 25.86 (Si^{-t}Bu), 24.53 (C-5), 21.57 (C-4), 20.24 (C-17Me), 18.14 (Si-C), 16.34 (C-11Me), 15.71 (C-19Me), 15.30 + 15.24 (B-C-CH₃?), 14.60 (C-28Me), 13.53 + 13.43 (B-C-C-CH₃?), 10.02 (C-25Me), -4.52 (SiMe), -4.73 (SiMe). FAB-MS (m/z): 1112, 1094, 1076; calc. for C₅₈H₁₀₂BNO₁₂Si: 1043.

Treatment of 16a (110mg, 0.1 mmol) with sodium perborate (45mg, 0.3 mmol) in 2ml of a 1:1 mixture of THF and water for 2h gave after chromatographic purification (toluene/ethyl acetate 2:1) 50mg (54%) 3a.

33-O-tert.Butyldimethylsilyl-22(S)-dihydro-FR-900520 (3b) was similarly prepared from 33-O-tert.butyldimethylsilyl-FR-900520 (14b) by reduction with tetramethylammonium-triacetoxyborohydride in 66% yield. ¹H-NMR (CDCl₃), two conformers (3:2), signals of the major conformer (cis-amide): δ 5.30 (s, H-26), 4.65 (d, J=5Hz, H-2), 4.53 (s, 10-OH), 4.42 (d,br, J=14Hz, H-6e), 3.62 (dd, J=1Hz, J=4.5Hz, H-14), 3.58 (m, H-15). ¹³C-NMR (CDCl₃), signals of the major conformer: δ 196.1 (C-9), 169.2 (C-1), 165.1 (C-8), 136.4 (C-19), 132.2 (C-28), 128.6 (C-29), 127.3 (C-20), 97.0 (C-10), 84.1 (C-32), 75.4 (C-15), 75.2 (C-33), 73.8 (C-13), 72.7 (C-14), 71.9 (C-22), 71.0 (C-24) 58.1 (OMe), 57.0 (OMe), 56.9 (C-2), 56.3 (OMe), 49.5 (C-18), 46.2 (C-21), 40.1 (C-25), 39.4 (C-6), 36.7 (C-23), 34.8 (C-30), 33.8 (C-34), 32.8 (C-16), 32.6 (C-12), 31.0 (C-35), 27.3 (C-3), 26.7 (C-17), 25.9 $(Si^{-t}Bu)$, 24.6 (C-5), 23.7 (C-36), 21.3 (C-4), 20.7 (17Me), 16.3 (11Me), 15.4 (19Me), 14.6 (C-27, 12.5 (C-37, 9.2)(25Me).

22(S)-Dihydro-FK-506 (6a)

a) From FK-506 with tetramethylammonium-triacetoxyborohydride. The same procedure as above can also be applied to reduce unsubstituted FK-506. 80mg crude product, containing small amounts of starting material and a more polar compound as side components (TLC, ethylacetate), was obtained from 80mg FK 506. However the purification of the product is much more tedious as compared to the silylated derivative. In one chromatographic purification step (ethylacetate) only 20mg (25%) pure **6a** could be isolated.

¹H-NMR (CDCl₃), two conformers (3:2), signals of the major conformer: δ 1.62 (H-23a), 1.25 (H-23b), 3.92 (H-24), 1.90 (H-25), 5.30 (H-26). ¹³C-NMR (CDCl₃): δ 195.9 (C-9), 169.3 (C-1), 164.9 and 165.4 (C-8), 137.4 and 136.9 (C-37), 136.3 and 135.7 (C-19), 126.2 and 126.8 (C-20), 97.0 and 98.4 (C-10), 71.7 and 70.9 (C-22), 43.9 and 44.1 (C-21), 36.8 and 36.9 (C-23).

b) From 33-0-tert.butyldimethylsilyl-22(S)-dihydro-FK-506 bydeprotection. 33-0-Tert.butyldimethylsilyl-22(S)-dihydro-FK-506 (**3a**) (276mg, 0.3 mmol) in 2ml acetonitrile was treated with 40% aqueous HF (20µl) for 1h at 10° C. The volatile components were evaporated in vacuum and the residue purified by chromatography (dichloromethane/ethanol 95:5) to give 241mg (87%) **6a**.

33-O-tert.Butyldimethylsilyl-22(S)-dihydro-FK-506-22,24-cyclic carbonate (4a) and 33-O-tert.butyldimethlysilyl-22(S)-dihydro-iso-FK-506 (5a)

33-O-tert.Butyldimethylsilyl-22(S)-dihydro-FK-506 (**3a**) (459mg, 0.5 mmol) was treated with carbonyldiimidazole (85mg, 0.5 mmol) in benzene (5ml) 12h at 50° C. The solvent was evaporated and the residue fractionated by chromatography (toluene/ethyl acetate 3:1) to give 165mg (35%) **4a** and

120mg (26%) 5a. 4a: 1 H-NMR (CDCl₃): δ 5.14 (H-26), 5.07 (H-20), 4.58 (H-22), 4.40 (H-24), 2.57 (H-21), 2.0 (H-25), 1.98 (H-23), 1.82 (H-23'). 13 C-NMR (CDCl₃): δ 196.2 (C-9), 169.1 (C-1), 164.9 (C-8), 138.2 (C-19), 135.7 (C-37), 131.0 (C-29), 129.9 (C-28), 123.4 (C-20) 117.1 (C-38), 97.0 (C-10), 84.1 (C-32), 77.8 (C-26), 75.4 (C-15), 75.0 (C-33), 73.6 (C-13), 72.7 (C-14), 56.3 (C-2), 49.5 (C-18), 42.9 (C-21), 39.8 (C-25), 39.7 (C-6), 36.4 (C-23), 36.1 (C-36), 34.9 + 34.8 (C-30, C-31), 33.9 (C-11), 33.7 (C-16), 32.8 (C-12), 30.8 (C-25), 26.1 (C-17), 21.2 (C-4), 20.0 (17Me), 18.1 (Si^tBu), 14.5 (C-27), 10.5 (25Me). FAB-MS (m/z): 946 (MH⁺), <u>928</u> (-H₂0), 702, 266. 5a: NMR data (CDCl₃) see table 1. FAB-MS (m/z): 902 (MH⁺-H₂0), 884 (-H₂0), 866 (-H₂0), 572, <u>266</u>.

33-O-tert.Butyldimethylsilyl-22(S)-dihydro-iso-FK-506 (5a)

A solution of 33-0-tert.butyldimethylsilyl-22(S)-dihydro-FK-506 (3a) (1g,1.09 mmol) and 4-dimethylaminopyridine (133mg,1.09 mmol) in 10ml DMF was stirred 7 days at 50° C. The solvent was evaporated in vacuum and the crude product purified by chromatography (toluene/ethyl acetate 1:1) to give 0.7g (70%) 5a. When imidazole was used instead of 4-dimethylamino-pyridine 5a was obtained in 62% yield after 7 days at 50° C.

33-0-tert.butyldimethylsilyl-22(S)-dihydro-iso-FR-900520 (5b)

The same procedure was used as for the preparation of 5a from 3a, but 33-O-tert.butyldimethylsilyl-22(S)-dihydro-FR-900520 reacted slower as compared to the FK 506 analogue: 47% product together with 47% starting material were isolated after 7 days and 62.5% product together with 22% starting material after 10 days stirring with 4-dimethylaminopyridine in DMF at 50° C. 1H-NMR (CDCl₃): δ 5.15-5.22 (m, H-24, H-29), 5.0 (d,br J=3Hz, H-2), 4.75 (d, J=10Hz, H-20), 4.44 (d,br J=13Hz, H-6e), 4.10 (s,br OH), 3.84 (d, J=6Hz, H-26), 3.66 (d, J=9Hz, H-14), 3.59 (dd, J=11Hz, J= 2.5Hz, H-15), 3.42 (s) + 3.39 (s) + 3.29 (s) (OCH₃), 0.90 (s, Si^tbutyl), 0.08 (s) + 0.07 (s) (Si-CH₃). FAB-MS (m/z): 890 (MH⁺-H₂O), 872 (-H₂O), 560, 266.

22(S)-Dihydro-iso-FK-506 (7a)

a) From 33-0-tert.butyldimethylsilyl-22(S)-dihydro-iso-FK-506 (5a).

33-O-tert.Butyldimethylsilyl-22(S)-dihydro-iso-FK-506 (5a) (400mg, 0.43 mmol) in 10ml THF was treated with 8ml 1N HCl 30min at room temp. The mixture was neutralized with aqueous NaHCO₃ and extracted with diethyl ether. The extract was dried (MgSO₄) and the solvent was evaporated to give 325mg 7a (94%). NMR data: comp. to 5a (tab.1). FAB-MS (m/z): 788 (MH⁺-H₂O), 770 (H₂O), 572, <u>266</u>.

b) From 22(S)-dihydro-FK-506 (6a) (qualitative). Reacting 22(S)-dihydro-FK-506 (20mg, 0.03 mmol) with 3mg 4-dimethylamino-pyridine in 0.5ml DMF at 50°C for 16 h. leads to a product (educt/product ~3:1) which was identified as 5a by TLC comparison.

22,26-Epoxy-22(S)-dihydro-iso-FK-506 (8a)

22 (S) -Dihydro-iso-FK-506 (120mg, 0.13mmol) and a few mg of p-toluenesulfonic acid in 2ml toluene were stirred for 24h at 60° C. The reaction mixture was filtered and the solvent was evaporated. Chromatography (ethylacetate) of the residue yielded 86mg (73%) 8a. ¹H-NMR (CDCl₃): δ 4.58 (dd, J=10.2, J=10.4Hz, H-25, 3.33 (m, H-22), 3.19 (d, J=10.2Hz, H-26), 4.58 (dd, J=10.4, J=5.4Hz, H-24), 1.68 (ddd, J=5.4, J=12.0, J=2Hz, H-23e), 1.35 (ddd, J=10.4, J=12.0, J= 10.0Hz, H-23a). FAB-MS (m/z): 788 (MH⁺), 770 (-H₂O), <u>5</u>72, 266.

22,33-0,0'-Bis-tert.butyldimethylsilyl-22(8)-dihydro-26-oxo-iso-FK-506 (11a) and 33-0-tert.butyldimethylsilyl-26-oxo-iso-FK-506 (10a)

33-O-tert.Butyldimethylsilyl-22(S)-dihydro-iso-FK-506 (5a) (254mg, 0.28 mmol) was treated with N-methyl-morpholine-N-oxide (71mg, 0.6 mmol) and a few mg tetrapropylammonium-perruthenate in 4ml dichloromethane in the presence of a 4A molsieve. After 1h stirring at room temp. the mixture was filtered and the solvent evaporated. The residue was purified by flash chromatography (toluene/ethyl acetate 4:1) and dissolved in 6ml dichloromethane. tert.Butyldimethylsilyltriflate (180mg, 0.75 mmol) and lutidine (90µl, 0.8 mmol) were added at 0° C and the mixture stirred overnight at RT. Routine workup gave 234mg crude product from which 135mg (47%) 11a and 38mg (15%) 33-0-tert.butyldimethylsilyl-26-oxo-iso-FK-506 (10a) were isolated by chromatographic separation (toluene/ethyl acetate 5:1).

11a ¹H-NMR (CDCl₃): δ 4.61 (H-2), 4.42 (H-6e), 3.11 (H-6a), 3.64 (d, J=9Hz, H-14), 3.59 (dd, J=11Hz, J=2.5Hz, H-15), 4.80 (d, J=9Hz, H-20), 5.07 (dd, J= 10Hz, J=2.5Hz, H-24), 6.24 (d, J=9Hz, H-29), 5.67 (ddt, J=17Hz, J=10Hz, J=7Hz, H-27), 4.97 (H-38tr), 4.91 (H-38cis), 3.40 + 3.38 + 3.28 (OCH₃). 10a ¹H-NMR (CDCl₃): δ 3.58 (d, J=9Hz, H-14), 2.93 (H-23a), 2.57 (H-23b) (J_{23a/23b}=17.5Hz, J_{23a/24}=9Hz, J_{23b/24}=2.5Hz), 5.55 (J_{24/25}, H-24), 3.70 (H-25), 1.07 (d, J=7Hz, 25-CH₃), 6.37 (d, J=9Hz, H-29).

33-O-tert.Butyldimethylsilyl-26-oxo-iso-FK-506 (10a) by oxidation of 5a with Dess-Martin reagent

Pyridine (32mg, 0.4mmol) and freshly prepared periodinane (84.8mg, 0.2mmol) were added to a solution of 33-O-tert.butyldimethylsilyl-22(S)dihydro-iso-FK-506 (5a) (138mg, 0.15mmol) in 10ml dichloromethane and the mixture was stirred for 15min at room temp. After cooling to 0°C 2ml of saturated aqueous Na_2SO_3 were added. The organic layer was separated, dried (MgSO₄) and the solvent was evaporated. Upon chromatography on silica gel (toluene/ethylacetate 4:1) 57mg (41%) 10a were obtained together with 60mg of a fraction which was a mixture of 10a and the 26-oxo-22-dihydro-derivative 9a.

33-O-tert.Butyldimethylsilyl-22(S)-dihydro-26-O-trimethylsilyl-iso-FK-506 (12a)

a) With trimethylsilyltriflate. Trimethylsilyltriflate (0.56g, 2.5 mmol) was slowly added at -70°C to a solution of 33-0-tert.butyldimethylsilyl-22(S)-dihydro iso-FK-506 (5a) (1.81g, 2mmol) and 2,6-lutidine (0.85g, 8 in 160ml dichloromethane and the mixture was stirred for further mmol) 30min. Aqueous NaHCO₂ was added and the organic layer separated, washed with water and brine, dried (MgSO,) and evaporated. Chromatography of the (toluene/ethylacetate 10:1 to 4:1) residue gave 349mg (17%) 22,26-0,0'-bis(trimethyl-silyl)-33-0-tert.butyldimethylsilyl-22(S)-dihydro-iso-FK-506 and 1.11g (57%) (12a). ¹H-NMR (CDCl₂): δ 5.71 (ddt, J=17Hz, J=10Hz, J=7Hz, H-37), 4.9-5.15 (5H: H-2, H-24, H-29, H-38tr, H-38cis), 4.80 (d, J=9Hz, H-20), 4.52 (H-6e), 3.96 (d, J=1Hz, 10-OH), 3.55-3.70 (3H: H-14, H-15, H-26), 3.44 + 3.39 + 3.29 (3 OCH₂).

b) With bis-(trimethylsilyl)-acetamide. 33-0-tert.Butyldimethylsilyl-

22(S)-dihydro-iso-FK-506 (5a) (322mg, 0.35 mmol) was treated with bis-(trimethylsilyl)-acetamide (142mg, 0.7 mmol) in 0.5ml toluene for 1 h at room temp. After quenching with water and extraction with toluene the organic solution was dried and the solvent evaporated. Upon chromatography of the residue (toluene/ethylacetate 4:1) 80mg (23%) 12a was obtained along with 75mg (22%) 33-0-tert.butyl-22(S)-dihydro-22-0-trimethylsilyliso-FK-506 and 160mg (43%) 22,26-0,0'-bis(trimethylsilyl)-33-0-tert.butyldimethylsily1-22(S)-dihydro-iso-FK-506. 33-0-tert.butyl-22(S)-dihydro-22-0-trimethylsilyl-iso-FK-506: ¹H-NMR (CDCl₂): δ 4.81 (H-2), 4.48 and 3.03 (H-6), 3.69 (H-14), 3.62 (H-15), 4.78 (H-20), 2.51 (H-21), 3.30, J_{22/23b}=11Hz, H-22), 1.69 (H-23a) and 1.38 (H-23b) $(J_{23a/23b}=13Hz, J_{23a/24}=10Hz), 5.13 (J_{24/25}=10Hz)$ 2.5Hz, H-24), 1.79 (J_{25/26}=6Hz), 3.87 (H-26), 5.20 (H-29), 2.97 (H-32), 0.90 (t-butyl), 0.17 (Me₃Si), 0.08 (Me₂Si).

33-O-tert.Butyldimethylsilyl-26-O-trimethylsilyl-iso-FK-506 (13a)

To a solution of 33-0-tert.butyldimethylsilyl-22(S)-dihydro-26-0-trimethylsilyl-iso-FK-506 (12a) (65mg, 0.06 mmol) and N-methyl-morpholine-N-oxide (about 40mg, 34 mmol) in 2ml dichloromethane were added a few mg of tetrapropylammonium-perruthenate as well as pulverized 4A molsieve and the mixture was stirred overnight at RT. Filtration, evaporation and chromatography (toluene/ethylacetate 6:1) yielded 45mg (69%) 13a. ¹H-NMR (CDCl₃): δ 4.67 (broad d, J=4Hz, H-2), 4.46 (broad d, J=13Hz, H-6e), 3.62 (d, J=9Hz, H-14), 3.55 (dd, J=11Hz, J=2.5Hz, H-15, 2.75 (H-23a) and 2.63 (H-23b) (J_{23a/23b}=17.5Hz, J_{23a/24}=7.5Hz, J_{23b/24}=5Hz), 5.20 (H-24), 3.93 (d, J=6Hz, H-26), 5.67 (ddt (J=17Hz, J=10Hz, J=7Hz, H-37), 3.43 + 3.39 + 3.29 (3 OCH₂). FAB-MS: 900 (MH⁺-Me₃SiOH), 882 (-H₂O), 718, 266, <u>226</u>.

Iso-FK-506 (2a) by deprotection of 13a

33-O-tert.Butyldimethylsilyl-26-O-trimethylsilyl-iso-FK-506 (13a) (28mg, 0.03mmol) in 1ml acetonitrile was treated with 0.5ml 1NHCl 30min at room temp. Saturated aqueous NaHCO₃ solution was added and the mixture extracted with ethyl ether. The ether solution was dried (MgSO₄), the solvents were evaporated and the residue was purified by chromatography (ethyl acetate) to give 20mg (88%) **2a**.

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Attempted rearrangement of 33-0-tert.butyldimethylsilyl-FK-506 or FK-506

a) With imidazole. After stirring a solution of 33-0-tert.butyldimethylsilyl-FK-506 (14a) (184mg) and imidazole (14mg) in 2ml DMF for 10 days at 50°C only unchanged starting material was recovered.

b) With tetraisopropoxytitanium. FK-506 (160mg, 0.2 mmol) was treated with tetraisopropoxytitanium (16ul, 0.05mmol) and 4A molsieve for 24h in dichloromethane (1ml) at reflux . According to TLC analysis the main component of the resulting reaction mixture is unchanged starting material. 2a is not detected.

Attempted intermolecular transesterification

33-O-tert.Butyldimethylsilyl-FR-900520 (14b) (200mg, 0.2mmol) was treated with tetraisopropoxytitanium (114µl, 0.4mmol) and methanol (0.1ml, 25 mmol) in 2ml THF at reflux for 3h. The reaction was quenched by addition of 1N HCl, the reaction mixture filtered through Celite and the solvent evaporated. The residue was purified by chromatography (toluene/ethyl acetate 2:1) to give 77mg (38%) benzilic acid rearrangement product 15b. ¹H-NMR (CDCl₃), two conformers (4:3): δ 3.83 + 3.63 (COOMe). ¹³C-NMR (CDCl₃), signals of the major conformer: δ 209.4 (C-22), 172.1 (C-8), 169.5 + 169.0 (168.4?) (C-1, C-9), 139.3 (C-19), 136.5 (C-29), 131.6 (C-28), 123.8 (C-20), 84.2 (C-32), 82.9 (C-10?), 79.2 (C-14), 77.7 (C-13), 75.2 (C-33), 68.3 (C-24).

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